

Application No.: 10/825,405

Docket No.: NY-NIAD 216-US2-DIV

REMARKS

Entry of the foregoing amendment is requested.

Applicants would like to draw the attention to two, non-prior art papers by the inventors, which support the claimed invention, i.e.:

M. Jacobson, et al., "Effect of myristyl nicotinate on retinoic acid therapy for facial photodamage," Exp. Dermatol., 16:927-935 (2007);

E. Jacobson, et al., "A topical lipophilic niacin derivative increases NAD, epidermal differentiation and barrier function in photodamaged skin," Exp. Dermatol., 16:490-499 (2007).

\* \* \*

Applicant believes no fee is due with this response. However, if a fee is due, please charge our Deposit Account No. 50-0624, under Order No. NY-NIAD 216-US2-DIV (10404746) from which the undersigned is authorized to draw.

Dated: January 8, 2008

Respectfully submitted,

By 

Norman D. Hanson

Registration No.: 30,946

FULBRIGHT & JAWORSKI L.L.P.

666 Fifth Avenue

New York, New York 10103

(212) 318-3000

(212) 318-3400 (Fax)

Attorney for Applicant

DOI:10.1111/j.1600-0625.2007.01535.x  
www.blackwellpublishing.com/EXP

Original Article

## Effect of myristyl nicotinate on retinoic acid therapy for facial photodamage

Myron K. Jacobson<sup>1,2</sup>, Hyuntae Kim<sup>1,2</sup>, W. Russell Coyle<sup>1,2</sup>, Moonsoon Kim<sup>1,2</sup>, Donna L. Coyle<sup>1,2</sup>, Ronald L. Rizer<sup>3</sup> and Elaine L. Jacobson<sup>1,2</sup>

<sup>1</sup>Department of Pharmacology & Toxicology, College of Pharmacy, and Arizona Cancer Center, University of Arizona, Tucson, AZ, USA;

<sup>2</sup>Niadene Development, Inc., Tucson, AZ, USA;

<sup>3</sup>Thomas Stephens & Associates, Inc., Colorado Springs, CO, USA

Correspondence: Myron K. Jacobson, PhD, Arizona Cancer Center, University of Arizona, 1515 N. Campbell Ave, Tucson, AZ 85724, USA.  
Tel.: +1 520 626 5953, Fax: +1 520 626 8567, e-mail: mjacobson@pharmacy.arizona.edu

Accepted for publication 11 July 2007

**Abstract.** Based on the hypothesis that skin barrier impairment is a contributor to side-effects associated with retinoic acid therapy, a double-blind, placebo-controlled pilot study examined the combined use of retinoic acid with myristyl nicotinate (MN), a lipophilic derivative of niacin that enhances skin barrier function, in female subjects with mild to moderate facial photodamage. The study involved a 1-month run-in period with placebo or MN prior to initiation of retinoic acid therapy for 3 months. Analysis of skin biopsies revealed that retinoic acid therapy resulted in stratum corneum thinning of approximately 25% ( $P = 0.006$  versus baseline) that was ameliorated by MN use ( $P < 0.005$ ). Therapy resulted in an increased rate of transepidermal water loss (TEWL) of approximately 45% ( $P = 0.001$  versus baseline) and use of MN protected against the increase in TEWL with the strongest protection provided by prior use of MN ( $P = 0.056$  versus

placebo). MN use reduced the incidence of side-effects of the therapy and again prior use provided the greatest reduction of side-effects. Subjects showed statistically significant clinical improvement ( $P < 0.05$  versus baseline) during the study; MN use did not interfere with any clinical improvement parameters and improved effects on temple laxity ( $P = 0.01$  versus placebo). Analysis of changes in epidermal thickness, Ki67-positive cells and intensity of lorcin staining demonstrated that MN either improved or did not interfere with retinoic acid efficacy. These results show that prior and concurrent use of MN can mitigate barrier impairment and improve the tolerability of retinoic acid therapy for facial photodamage without interfering with efficacy.

**Key words:** facial photodamage – myristyl nicotinate – retinoic acid – skin barrier function

Please cite this paper as: Effect of myristyl nicotinate on retinoic acid therapy for facial photodamage. *Experimental Dermatology* 2007; 16: 927–935.

### Introduction

Retinoids, the natural metabolites of vitamin A and synthetic vitamin A analogues, are important regulators of skin function (1). All-*trans*-retinoic acid (vitamin A acid), the major naturally occurring biologically active retinoid, has been a focus of research into topical treatments for photodamaged skin for many years. Kligman et al. reported that retinoic acid could produce smoother, less wrinkled and less pigmented skin after a few months of treatment (2). Retinoids have prominent pharmacological effects in both epidermal and dermal compartments (3–5). In the epidermis of chronically photodamaged skin, long-term topical retinoid therapy results in dose-dependent increases in epidermal and granular layer thickness, stratum corneum compaction, decreased melanin content and improvement of epidermal atypia

(1,5–7). Retinoids induce proliferation in keratinocytes, presumably mediated by epidermal growth factor receptor activation resulting in epidermal hyperplasia (8). Retinoic acid-induced expression of keratins K6, K16 and K17, which are commonly expressed in the hyperproliferative epidermis, indicates that retinoids increase cell proliferation in the basal and/or lower spinous layers of the epidermis (9). Retinoids also can lighten hyperpigmented skin, reduce tyrosinase activity in cultured melanocytic cells (10,11), inhibit proliferation and lipid synthesis, and alter keratin expression in cultured human sebocytes (12). Dermal effects include increased fibroblast proliferation (4), increased collagen production (13) and reduced extracellular matrix degradation (1). Prolonged use of retinoic acid significantly increases collagen matrix deposition in dermal repair zones apparently responsible for the wrinkle reduction that accompanies retinoic acid treatment of photodamaged skin (5,11).

While retinoic acid provides multiple benefits to photodamaged skin (11), it is frequently accompanied by

**Abbreviations:** MN, myristyl nicotinate; TEWL, transepidermal water loss.

Jacobson et al.

significant skin irritation that limits compliance with therapy (14). The most commonly reported retinoic acid treatment-related adverse effects are irritation, dryness, peeling, erythema and a sensation of burning on the skin (14). The mechanisms leading to retinoid side-effects are still incompletely understood, but retinoic acid therapy is known to impair barrier function as assessed by transepidermal water loss (TEWL) measurements (15). Barrier impairment has been attributed to retinoid-induced epidermal hyperplasia (16) and to alteration of the terminal differentiation programme (1). Erythema, which reflects the production of epidermal cytokines such as interleukin-1, may result from retinoid-stimulated keratinocyte proliferation directly or as a consequence of epidermal barrier impairment (17,18). Retinoid-induced stratum corneum thinning, often referred to as compaction (6,7), is likely related to barrier impairment as stratum corneum thickness is a major determinant of barrier function (19,20).

Myristyl nicotinate (MN), a niacin derivative developed for optimal topical delivery of nicotinic acid to skin (21,22), has been shown to enhance epidermal differentiation in photodamaged skin, resulting in increased stratum corneum and epidermal thickness and enhanced barrier function (22). Based on the hypothesis that barrier impairment contributes to the side-effects of retinoic acid therapy, we report here the results of a double-blinded study simultaneously using a placebo or MN-containing skin cream with retinoic acid therapy in subjects with mild to moderate facial photodamage. The results demonstrate that concomitant use of MN mitigates retinoic-acid-induced loss of barrier function and improves tolerability without interfering with the efficacy of retinoic acid. Finally, we report that 1-month use of MN prior to initiation of retinoic acid therapy provides additional barrier protection and tolerability of retinoic acid without interfering with, and in some cases, improving efficacy.

## Methods

This study was a 12-week randomized, double-blinded, placebo-controlled evaluation of the effects of a 5% MN formulation on surrogate markers of barrier function, clinical and sensory irritation, and clinical efficacy associated with retinoic acid use. The study was conducted (July 2005 to December 2005) by a contract research organization, Thomas J. Stephens & Associates (Colorado Research Center, Colorado Springs, CO, USA). Healthy adult female subjects between the ages of 30 and 60, with a score of I to IV on the Fitzpatrick Skin Classification (23), mild to moderate photodamaged skin as defined by a modified Glogau Classification of I to II (24), and presence of dyschromia on the face as determined by a Woods light visual scan, were eligible for the study. The mean age of the study

population was  $43.4 \pm 6.96$  years and included 86.7% Caucasian, 8.3% Hispanic, 1.7% Asian and 3.3% other ethnicities. Subjects were randomly assigned to one of three groups of 20 subjects each [group 1 received placebo for 1 month prior to initiation of retinoic acid therapy (baseline), then placebo and retinoic acid (0.025%) from baseline to 12 weeks (P/P + RA); group 2 received placebo for 1 month, then MN (5%) and retinoic acid (0.025%) from baseline to 12 weeks (P/MN + RA); and group 3 received MN (5%) for 1 month, then MN (5%) and retinoic acid (0.025%) from baseline to 12 weeks (MN/MN + RA)]. The dose of retinoic acid was 0.1 g in all cases. This dose of retinoic acid was chosen for this study as the subjects had mild to moderate photodamage. Subjects also were provided with Cetaphil® liquid cleanser and Neutrogena Ultra Sheer SPF 30 sunscreen to use for facial cleansing and sun protection during the entire course of the study. Subjects applied the assigned test moisturizers at 2.5 mg/cm<sup>2</sup> [MN (5%) or a placebo that contained myristyl myristate in place of MN] to the entire face twice a day after cleansing. During the usage phase of the study, subjects applied the retinoic acid (0.025%) to the face after test moisturizer application once a day in the evening.

The study was conducted in accordance with the applicable Good Clinical Practice regulations and guidelines and Institutional Review Board regulations. All subjects were required to read and sign an IRB-approved Informed Consent Form. The sample size was determined empirically. Each subject who qualified for enrollment was assigned a subject number that was used in all documentation. The numbers were unique and were assigned in ascending order using a computer-generated randomization schedule developed by the contract research organization. The numbers were concealed until after the intervention was assigned. The contract research organization enrolled and assigned subjects to groups. All participants, those administering the interventions, and those assessing the outcomes were blinded to group assignment. Analyses were not by 'intention-to-treat' but only on those who completed the study. Analyses were restricted to subjects with complete data sets; thus, the *n* for some measures differs slightly because of missing data points for a given individual.

## Punch biopsies

A board-certified dermatologist collected a 2-mm punch biopsy from the right or left side of the face for 10 randomly selected subjects from each group at baseline and after 12 weeks of treatment. The punch biopsies were formalin-fixed, embedded in paraffin, cut into 5 µm cross-sections, mounted on slides and stained with haematoxylin-eosin (H&E). Tissue microarrays were generated using 1.5-mm cores from formalin-fixed, paraffin-embedded skin punch biopsies by the Translational Genomics Research Institute

Tissue Microarray Core Facility (TGEN, Phoenix, AZ, USA). Thirty 4- $\mu$ m slices from each array were cut, mounted onto glass slides and immunohistochemistry was performed.

#### TEWL measurements

Subjects were required to equilibrate to ambient conditions for at least 20 min and were maintained between 66 and 72°F and a relative humidity between 15% and 55%. A Dermalab instrument was used to measure TEWL at two points above the skin surface, and the rate of water loss was calculated. Each TEWL measurement was averaged over a 1-min measurement period.

#### Image analyses

Histological images were taken of H&E-stained cross-sections with an Olympus inverted stage microscope using a 10 $\times$  by 0.45 Apochromat objective and a Nikon digital CCD camera. ImageJ image analysis software (NIH) was used to examine the images and perform measurements. Suprapapillary epidermal thickness (as measured from the top of the dermal papilla to the top of the granular layer) and stratum corneum thickness (as measured from the top of the granular layer to the top of the stratum corneum) were measured. For each specimen, five different sites were measured and the average was calculated.

#### Immunohistochemistry analysis

Immunohistochemistry was performed using the Discovery XT autostainer by Ventana Medical Systems, Inc. (Tucson, AZ, USA). Diaminobenzidine detection solutions, the Ki67 antibody and the haematoxylin counter stain are proprietary reagents supplied by Ventana Medical Systems, Inc. for use on human tissues. Loricrin polyclonal antibody (PRB-145P; Covance Research Products, Berkeley, CA, USA) was used at a 1:100 dilution. Histological images were taken of Ki67 and loricrin-stained cross-sections with an Olympus inverted stage microscope using a 10 $\times$  by 0.45 Apochromat objective and a Nikon digital CCD camera. After acquisition with a digital camera, the image files were analysed using Photoshop (Adobe, San Jose, CA, USA). For the Ki67 analyses, a line was drawn on the electronic images to demarcate the epidermis in two equal sections between the basal layer and the stratum corneum. Stained cells in the lower half of the epidermis were marked with a red dot while unstained cells were marked with a green dot. Proliferation index was calculated as the total number of stained cells divided by the total number of cells in the section. For quantitative analyses of loricrin staining, the method of Matkowskyj et al. was used (25). Briefly, the 'Magic Wand tool' of Photoshop was used to select the entire stained region of the histological image. A threshold tolerance value of 30 was used for this tool to capture all pixels falling within the threshold parameter selected, which was

removed from the original image, placed in a new non-compressed TIFF file, and saved as the stained image. The original minus the stained region was saved as another non-compressed TIFF file as the unstained image. The amount of antibody staining was quantified using the TIFFalyzer program (University of Illinois Medical Center) outputting a RGB value between 0 (black) and 255 (white). Each deconstructed TIFF image, stained and unstained, was processed and an RGB value obtained. The final image energy was calculated by subtracting the stained image value from its unstained counterpart.

#### Clinical grading of tolerability and efficacy

Subjects were clinically graded on the right and/or left side of the face for efficacy/performance parameters and irritation/safety parameters at baseline and weeks 2, 4, 8 and 12. For tolerability assessment, parameters of scaling/peeling and degree of erythema were graded on a 3-point scoring system. The graders also questioned each subject at each scheduled visit and recorded the occurrence of tightness/dryness, stinging, burning and tingling side-effects. Clinical assessment of efficacy was assessed on a 5-point scale of five parameters associated with efficacy of retinoic acid therapy for facial photodamage that included dyschromia, fine lines, shallow wrinkles, tactile roughness and temple laxity.

#### Subject self-assessment of efficacy

Self-assessment questionnaires were administered at the completion of the study that requested study subjects to respond to questions with one of five choices (strongly agree, agree, neither agree nor disagree, disagree, strongly disagree) concerning their perception of a decrease in signs of ageing, disappearance of fine lines, increase in smoothness/softness, improvement in skin radiance and increase in healthy appearance of the skin.

#### Inclusion criteria

Eligible subjects were females between the ages of 30 and 60 who were in generally good health as determined by a health assessment questionnaire. Subjects were required to be willing to avoid direct sun exposure and the use of tanning beds for the duration of the study.

#### Exclusion criteria

Individuals were ineligible for this study if, in the opinion of the investigator, they had known allergies or sensitivities to products that may have influenced the study; exhibited any skin disorders on the test areas of the face that may have influenced the study; had known medical conditions, such as diabetes, that could affect wound healing; or were using medications that might have influenced the study (e.g. prescription strength steroids, prescription strength anti-inflammatory drugs or topical medications). Subjects

Jacobson et al.

were ineligible if they were pregnant or nursing. Other exclusion criteria included hypertension or uncontrolled metabolic disease, atopic diseases such as asthma, atopic dermatitis of the face, arms or hands; known sensitivity to alpha- and beta-hydroxyacids; use of products containing hydroxyacids or retinoids such as Retin-A®, Renova®, microdermabrasion treatment or routinely used alpha-hydroxy-acids, beta-hydroxy-acid or poly-hydroxy-acid products within 1 month of the study start; routine use of skin lightening products within 1 month of study start; had ablative and non-ablative laser treatments as well as Thermanage treatments on the face or arms; had a 'lunchtime' facial peel within the last year or; currently enrolled on another facial usage study.

## Results

### Myristyl nicotinate prevents retinoic acid-associated stratum corneum thinning

Haematoxylin-eosin stained sections from periorbital biopsy samples obtained from study subjects at baseline and 12 weeks were used to determine stratum corneum thickness (Fig. 1). At baseline, the mean stratum corneum thickness of the P/MN + RA group was slightly higher than the P/P + RA group, although the difference was not statistically significant. The mean thickness of the MN/MN + RA group at baseline, which had been treated for 1 month with 5% MN, was approximately 11% higher

than the other two groups, although the difference did not reach statistical significance at  $P < 0.05$ . However, previous studies have shown that treatment of photodamaged skin with 5% MN for 3 months results in an increase in stratum corneum thickness of more than 50% (22), and thus the trend towards a higher mean value in MN/MN + RA group compared with that of the other groups agrees with the known effects of MN on photodamaged skin. During the 12 weeks of retinoic acid therapy, the P/P + RA group experienced a reduction in stratum corneum thickness of approximately 24% ( $P = 0.006$  versus baseline), but concurrent initiation of MN use with retinoic acid therapy did not result in stratum corneum thinning. The difference in stratum corneum thickness between the P/P + RA and P/MN + RA groups at 12 weeks of therapy was highly statistically significant ( $P = 0.005$ ), even though the  $n$  in this pilot study was only 7. Stratum corneum thickness of the MN/MN + RA group decreased by approximately 2% (not statistically significant), but the difference in stratum corneum thickness between this group and the P/P + RA group at 12 weeks was highly statistically significant ( $P = 0.003$ ). These data show that use of MN mitigates stratum corneum thinning associated with retinoic acid therapy.

### Myristyl nicotinate reduces retinoic acid-associated barrier impairment as assessed by TEWL measurements

Transepidermal water loss rates provide a non-invasive assessment of relative barrier function (20,26,27). Thus, TEWL measurements taken from the faces of study subjects were used as a surrogate marker of barrier function to compare placebo- and MN-treated groups. Figure 2 (panel a) shows the change in TEWL on the right cheek from baseline to 12 weeks for each group. The rates of TEWL increased in the P/P + RA group by approximately 45%, a value that was highly statistically significant ( $P < 0.0001$ ) even though TEWL measurements made on the face have a documented high level of variability (19). The mean rates of TEWL also increased in the P/MN + RA and MN/MN + RA groups, although the changes from baseline for these groups were not statistically significant. The difference between the P/P + RA and MN/MN + RA groups at 12 weeks nearly reached statistical significance ( $P = 0.056$ ). Figure 2 (panel b) shows the time course of changes from baseline for the three groups. The rates of TEWL for the P/P + RA and P/MN + RA groups were similar at 2 weeks of retinoic acid therapy, but the TEWL values for the P/MN + RA group were lower thereafter. In contrast, the TEWL values for the MN/MN + RA group were consistently lower than the other groups' at all time points following initiation of therapy. These results indicate that concurrent use of MN mitigates barrier impairment and that prior plus concurrent use provides greater barrier protection than concurrent use alone.

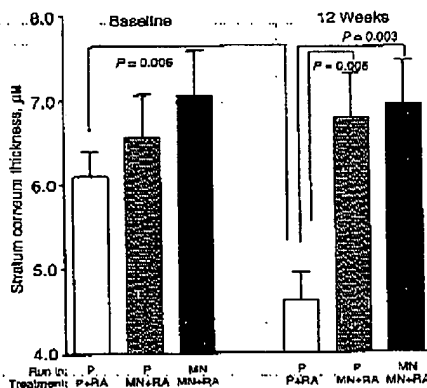
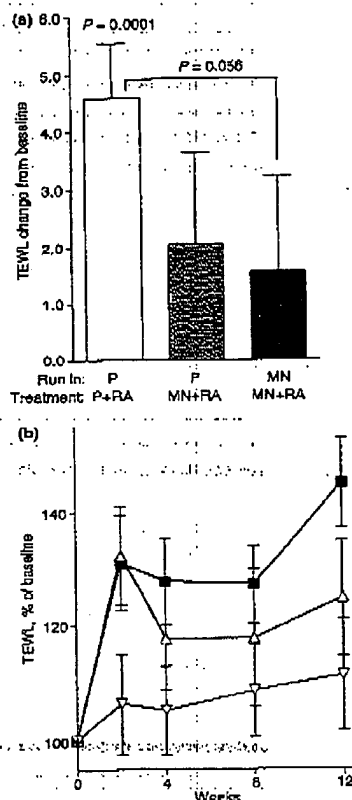


Figure 1. Effect of myristyl nicotinate (MN) on stratum corneum thickness during retinoic acid therapy. Biopsy samples from the periorbital regions were obtained at baseline and 12 weeks, haematoxylin-eosin (H&E) stained, and analysed for stratum corneum thickness ( $n = 7$ ). Open bars represent the P/P group (P/P + RA), grey bars represent the P/MN group (P/MN + RA), and black bars represent the MN/MN group (MN/MN + RA). P = placebo. The letters in parenthesis before the slash describe the 30 day run in cream and those after the slash describe the 3 month treatment creams, placebo or MN+ retinoic acid, starting at baseline. Error bars depict SEM. The P-values shown were derived from two-tailed Student's  $t$ -tests.



**Figure 2.** Effect of myristyl nicotinate (MN) on rates of transepidermal water loss (TEWL) during retinoic acid therapy. Rates of TEWL were determined on the right cheeks of study subjects as described in methods. The left panel shows the mean change in rates of TEWL from baseline to 12 weeks. Open bars represent the P/P group (P/P + RA) ( $n = 20$ ), grey bars represent the P/MN group (P/MN + RA) ( $n = 20$ ), and black bars represent the MN/MN group (MN/MN + RA) ( $n = 17$ ). P=placebo. The letters in parenthesis before the slash describe the 30 day run in cream and those after the slash describe the 3 month treatment creams, placebo or MN+ retinoic acid, starting at base line. The P-values shown were derived from unpaired, two-tailed Student's *t*-tests. The right panel shows the per cent change in TEWL from baseline for the P/P + RA (squares), P/MN + RA (triangles), and MN/MN + RA (inverted triangles) groups. Error bars depict SEM.

### Myristyl nicotinate improves the tolerability of retinoic acid therapy

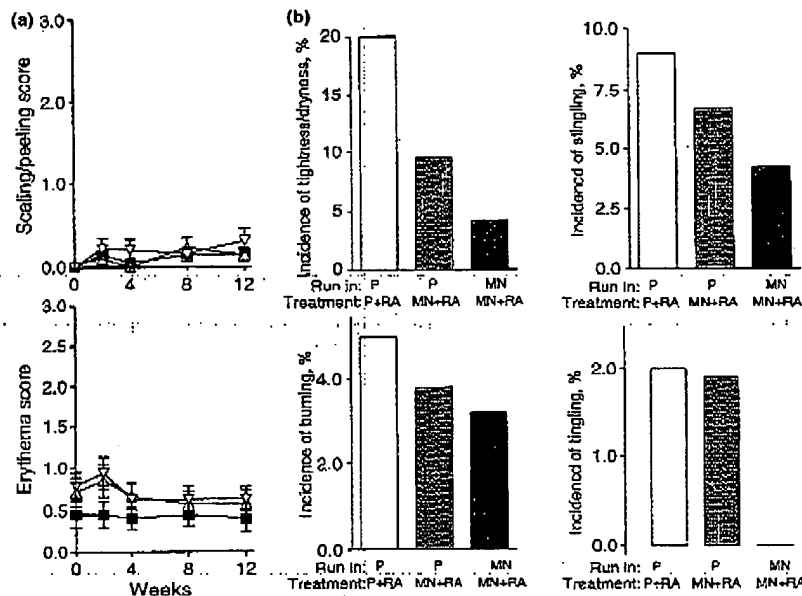
Expert clinical grading assessed the tolerability of retinoic acid therapy. The parameters of tolerability such as scaling/peeling and degree of erythema were graded on a 3-point scoring system. The frequency of less severe parameters of tolerability typical of retinoic acid therapy that included tightness/dryness, stinging, burning and tingling also were recorded by the clinical graders. The degree of scaling/peeling was very low in all groups, remaining

below 0.3 on the 3-point scale, and the degree of erythema also was relatively low as all scores were at 1.0 or below on the 3-point scale (Fig. 3, panel a), indicating an overall high degree of tolerance of the 0.025% concentration of retinoic acid when used with a moisturizer twice a day prior to and during therapy. There were no statistically significant differences between the placebo and MN groups in either parameter, although the grading of erythema was consistently slightly higher in the MN-treated subjects. Despite the low levels of scaling/peeling or erythema, a significant frequency of less severe but commonly encountered side-effects of retinoic acid was observed in this study (Fig. 3, panel b). For these tolerability parameters, a consistent pattern was observed as concurrent use of MN with retinoic acid decreased the frequency of tightness/dryness, stinging and burning, while prior and concurrent MN use further reduced the frequency of each of these parameters. Although the frequency of tingling reported was quite low (2%), the incidence of this side-effect was reduced to zero for the MN/MN + RA group. In addition to the clinical grading, study subjects completed self-assessment questionnaires that solicited information related to tolerability of the therapy. These self-assessments paralleled the clinical grading in all cases where the same parameter was assessed. Additionally, the study subjects reported a decreased frequency of comedones in the MN/MN + RA group. In total, the results show that use of MN improved the tolerability of retinoic acid therapy.

### Myristyl nicotinate does not interfere with and in some cases improves the efficacy of retinoic acid therapy as assessed by clinical grading and patient self-assessment

Expert clinical grading and patient self-assessment were used to assess the effect of MN on the efficacy of retinoic acid therapy on visible clinical parameters. Clinical grading involved evaluation of dyschromia, fine lines, shallow wrinkles, tactile roughness and temple laxity as a function of treatment time (Fig. 4). Despite some differences in the degree of initial photodamage between the groups, similar rates of improvement for all three groups were observed for each of the parameters evaluated. The improvements were statistically significant relative to baseline ( $P < 0.05$ ) at many of the time points. For tactile roughness, the MN/MN + RA group consistently showed greater improvement from weeks 4 to 12 compared with the P/P + RA group, although the difference did not reach statistical significance at  $P < 0.05$ . Grading of temple laxity showed a statistically significant greater improvement at 12 weeks ( $P = 0.01$ ) in the MN/MN + RA group compared with the P/P + RA group and a trend for greater improvement in the P/MN + RA compared with P/P + RA was observed that did not reach statistical significance at  $P < 0.05$ .

Jacobson et al.



**Figure 3.** Effect of myristyl nicotinate (MN) on side-effects associated with retinoic acid therapy as assessed by clinical grading. (Panel a) Effect of myristyl nicotinate on the severity of scaling/peeling and erythema as assessed by clinical grading. The degree of severity was assessed on a 3-point clinical scale and mean values were determined for the placebo/placebo (squares) ( $n = 20$ ), placebo/MN (triangles) ( $n = 21$ ), and MN/MN (inverted triangles) ( $n = 19$ ) groups. Error bars represent SEM. (Panel b) The incidence of tightness/dryness, stinging, burning and tingling side-effects. Open bars represent the P/P group (P/P + RA) ( $n = 20$ ), grey bars represent the P/MN group (P/MN + RA) ( $n = 21$ ), and black bars represent the MN/MN group (MN/MN + RA) ( $n = 19$ ). P = placebo. The letters in parenthesis before the slash describe the 30 day run-in cream and those after the slash describe the 3 month treatment creams, placebo or MN + retinoic acid, starting at base line. Incidence rates were determined from clinical grading at 2, 4, 8 and 12 weeks.

Patient self-assessment of efficacy groups using MN rated efficacy higher than subjects in the P/P + RA group in all categories examined. These results indicate that concurrent or prior and concurrent use of MN does not interfere with retinoic acid efficacy and that MN use can result in improved efficacy in some cases.

#### Myristyl nicotinate does not interfere with efficacy of retinoic acid therapy as assessed by analysis of skin biopsies

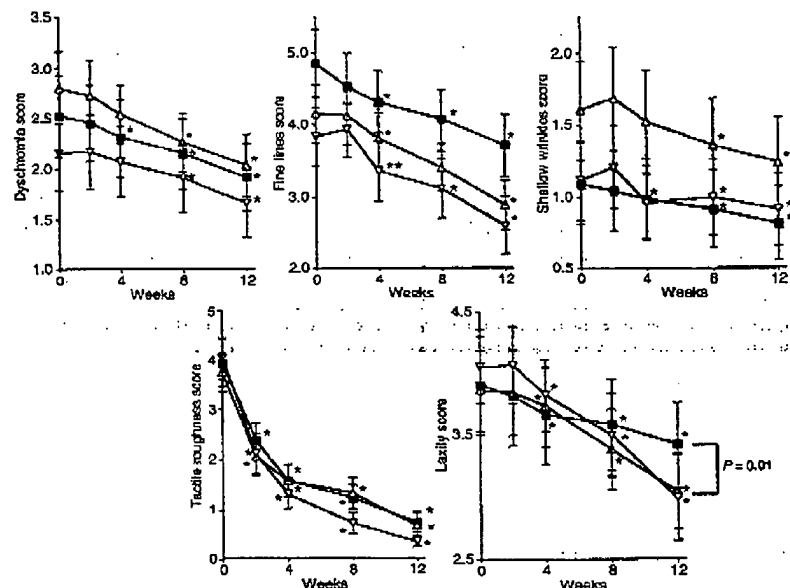
Retinoid therapy is associated with an initial decrease in epidermal thickness and then after approximately 6 months of therapy an increase in epidermal thickness (1,6,7). Changes in epidermal thickness in each of the groups over the 12-week course of the retinoic acid therapy were assessed (Fig. 5). The mean values for the P/P + RA, P/MN + RA, and MN/MN + RA groups at the baseline were 37.9, 38.8 and 39.3  $\mu\text{m}$ , respectively, which were not statistically significantly different even though the use of MN for 30 days showed a trend towards increased epidermal thickness, which is known to occur with an MN treatment over a longer period of time (22). The mean epidermal thickness of the group receiving retinoic acid and the placebo cream decreased by approximately 5% over the 12-week study, likely due to the limited duration of the study. The epidermal thickness of the group concurrently receiving MN increased by approximately 3%, and the group receiving MN prior to and concurrently with retinoic acid increased by approximately 10%. The difference between the P/P + RA and MN/MN + RA groups at

12 weeks was statistically significant ( $P = 0.0007$ ), while the difference between P/P + RA and P/MN + RA groups showed a trend but did not reach statistical significance at  $P < 0.05$ . The difference between the P/MN + RA and MN/MN + RA groups at 12 weeks also reached statistical significance ( $P = 0.05$ ). The results of epidermal thickness determinations support the possibility that MN use accelerates the efficacy of retinoic acid therapy.

The effects of MN on the frequency and localization of Ki67 as a marker of proliferation and on the intensity of loricrin staining as a marker of differentiation were assessed by immunohistochemistry. In all cases, Ki67-positive cells were located in the lower half of the epidermis and there was no effect of the 1-month run-in period with MN in comparison with the placebo on the percentage of Ki67-positive cells (data not shown). Retinoic acid therapy increased the frequency of Ki67-positive cells by 9–19%, consistent with a previous study (28), but there were no statistically significant differences between groups using placebo or MN formulations (data not shown). The 1-month use of MN resulted in a statistically significant ( $P = 0.01$ ) increase in the intensity of loricrin (Fig. 6, panel a). In panels b, c, and d of Fig. 6 loricrin staining intensity of each of the three groups was compared at the initiation of retinoic acid therapy (baseline) and following 12 weeks. In each case, a statistically significant increase in staining was observed. These results indicate that both MN and retinoic acid stimulate increased loricrin expression and that MN does not interfere with the ability of retinoic acid to stimulate loricrin expression.

## Myristyl nicotinate and retinoic acid therapy

**Figure 4.** Effect of myristyl nicotinate (MN) on efficacy of retinoic acid therapy as assessed by clinical grading. Clinical assessment on a 5-point scale of five parameters associated with efficacy of retinoic acid therapy for facial photodamage over the course of the study was completed as described in 'Methods'. Mean values were determined for the P/P + RA (squares) ( $n = 20$ ), P/MN + RA (triangles) ( $n = 21$ ), and MN/MN + RA (inverted triangles) ( $n = 19$ ) groups. The letters before the slash describes the 30 day run in cream and those after the slash represent the 3 month treatment period creams, placebo or MN + retinoic acid, starting at base line. Error bars represent SEM; data points marked by \* indicates  $P < 0.05$  versus baseline using paired, two-tailed Student's  $t$ -tests. The  $P$ -value shown in the lower right panel was derived from an analysis of variance (ANOVA), with paired comparisons (Fisher's LSD) between the P/P + RA and MN/MN + RA groups.



## Discussion

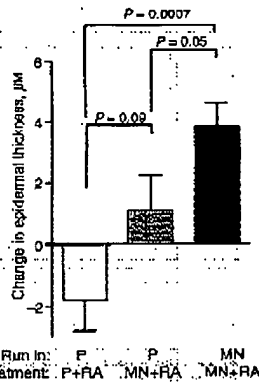
The correlation between stratum corneum thinning (Fig. 1) and increased rates of TEWL (Fig. 2) support the hypothesis that stratum corneum compaction is a factor in barrier impairment associated with retinoic acid therapy. In the study reported here, changes in stratum corneum thickness have been determined in paraffin-embedded samples subjected to the same conditions of preparation as the placebo samples. While prior studies have shown that the thickness of the stratum corneum is less in paraffin sections compared with frozen sections (29), paraffin sections have been used to assess stratum corneum morphology and changes in thickness in numerous studies (7,30–32). Previous studies of the effects of MN on photodamaged skin have shown that this agent stimulates increases in both epidermal and stratum corneum thickness and results in increased barrier function as evidenced by decreased rates of TEWL (22). Our results here show that concurrent and prior plus concurrent use of MN reduces stratum corneum thinning (Fig. 1) and reduces the increase in rates of TEWL in subjects on retinoic acid therapy (Fig. 2). A possible link between barrier impairment and the irritation potential of retinoic acid is supported by studies of other dermatology conditions that link barrier impairment with high irritation and/or inflammation potential (33–35). The results presented in this study support this link as the ability of MN to reduce stratum corneum thinning and reduce barrier impairment was coincident with a reduced frequency of tightness/dryness, stinging, burning and tingling side-

effects (Fig. 3, left panel). It is interesting that 1-month pretreatment with MN showed a pattern of less side-effects than did use initiated concurrent with initiation of retinoic acid therapy (Fig. 3) and that the improved tolerability was coincident with positive effects of pretreatment on barrier impairment as assessed by TEWL measurements (Fig. 2). As previous studies have observed that MN-related increases in stratum corneum and epidermal thickness and decreases in rates of TEWL in photodamaged skin are progressive over a period of several months (22), additional studies on optimizing duration of pretreatment with MN on retinoid therapy for skin photodamage and possibly even acne are warranted. The feasibility of such studies is suggested by the excellent tolerability profile of formulations containing MN (22). It was also noted that a slightly higher grading of erythema in subjects using MN was observed. Prior studies of actinic skin damage have shown that MN treatment results in changes in skin tone (E.L. Jacobson, unpublished data), and whether this or other mechanisms led to the slightly higher erythema scoring also needs to be examined.

A key question addressed in this study was whether MN would negatively affect the efficacy of retinoic acid therapy. The results of clinical grading (Fig. 4) and of patient self-assessments argue that this is not the case. Similar rates of improvement were observed between placebo and MN groups, and for temple laxity at 12 weeks, a statistically significantly increased efficacy was observed with the use of MN. The epidermal thickness measurements reported here (Fig. 5) also indicate that MN use does not interfere with



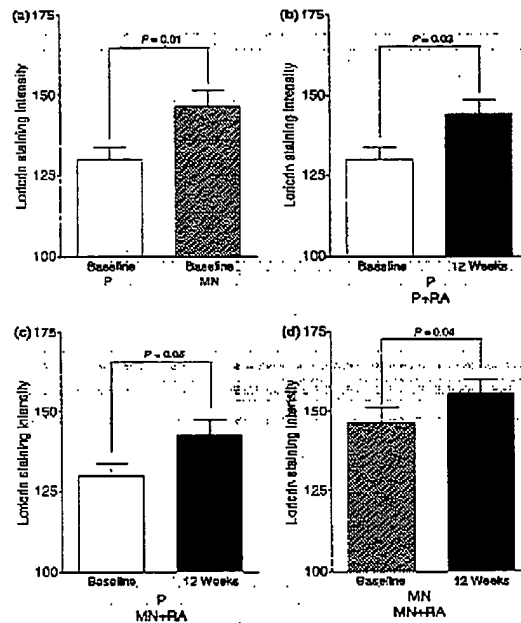
Jacobson et al.



**Figure 5.** Effect of myristyl nicotinate (MN) on epidermal thickness during retinoic acid therapy. Biopsy samples from the periorbital regions were obtained at baseline and 12 weeks, H&E stained, and analysed for epidermal thickness. The number of biopsy samples analysed was 7 in each group. Open bars represent the (P/P + RA) group, grey bars represent the (P/MN + RA) group, and black bars represent the (MN/MN + RA) group. The letters in parenthesis before the slash describe the 30 day run in cream and those after the slash describe the 3 month treatment creams, placebo or MN+ retinoic acid, starting at base line. P = placebo; RA = retinoic acid. Error bars depict SEM. The *P*-values shown were derived from unpaired, two-tailed Student's *t*-tests.

efficacy and are consistent with the possibility that MN accelerates the efficacy of retinoic acid, although it is also possible that the differences observed reflect the known property of MN to enhance epidermal thickness in photo-damaged skin (22). The intensity of loricrin staining was used as a marker of differentiation and the results show that MN alone stimulates loricrin expression (Fig. 5, panel a), that retinoic acid increases loricrin expression (Fig. 5, panel b), consistent with previous reports (32), and that MN does not interfere with increased loricrin expression during retinoic acid treatment (Fig. 5, panels c and d). In total, our results do not indicate any deleterious effect of MN on retinoic acid therapy.

The progression of actinic skin damage involves increased proliferation and de-differentiation of clones of damaged cells in the epidermis that can progress to actinic keratosis lesions and non-melanoma skin cancer (21). The ability of retinoic acid to reduce atypical cells in actinic skin damage suggests that it may provide skin cancer prevention benefit (11). The ability of oral retinoic acid to reduce skin cancer risk has been reported (36), providing additional support for a possible role in skin cancer prevention. The ability of MN to promote epidermal differentiation as evidenced by increases in epidermal and stratum corneum thickness (22) and loricrin expression (Fig. 6) raises the hypothesis that this agent also may limit the progression of early-stage actinic skin damage to actinic keratoses and non-melanoma skin cancer. Other studies



**Figure 6.** Effect of myristyl nicotinate (MN) on loricrin staining prior to and during retinoic acid therapy. The intensity of loricrin staining was determined as described in 'Methods'. (Panel a) Compares relative staining intensity following the 1-month run-in period with placebo (open bar, *n* = 14) or MN (stippled bar, *n* = 9) formulations. The relative staining intensity at baseline (open bar, *n* = 14) and 12 weeks (solid bar, *n* = 7) is shown for the (P/P + RA) group in panel b. (30 day run in cream/3 month treatment creams) P=placebo; RA=retinoic acid. (Panel c) Compares intensity at baseline (open bar, *n* = 14) and 12 weeks (solid bar, *n* = 7) for the (P/MN + RA) group. (Panel d) Compares intensity at baseline (stippled bar, *n* = 9) and 12 weeks (solid bar, *n* = 9) for the (MN/MN + RA) group. The *P*-values were determined using an unpaired Student's *t*-test for panel a and a paired two-sided Student's *t*-test for panels b, c, and d.

also support the concept that optimal niacin status may limit carcinogenesis via enhancing genomic integrity (37,38). Indeed, the possibility that the combined use of retinoic acid and MN may effectively limit progression of actinic skin damage deserves consideration.

## Acknowledgments

These studies were funded in part by Niadyne, Inc., and data analyses were funded by NIH grant R44 CA-90085 and P01 CA027502. Histological processing of biopsy samples was done by the TACMASS Core at the Arizona Cancer Center supported by NIH grant CA 23074. We thank Brett Bourguet for Ki67 quantification. MKJ and ELJ are principals in Niadyne, Inc., whose sponsored research is managed in accordance with the University of Arizona conflict-of-interest policies.

## References

- 1 Fisher G J, Voorhees J J. Molecular mechanisms of retinoid actions in skin. *FASEB J* 1996; 10: 1002-1013.
- 2 Kligman A M, Grove G L, Hirose R, Leyden J J. Topical tretinoin for photoaged skin. *J Am Acad Dermatol* 1986; 15: 836-859.
- 3 Gendimenico G J, Mezick J A. Pharmacological effects of retinoids on skin cells. *Skin Pharmacol* 1993; 6 (Suppl. 1): 24-34.
- 4 Varani J, Warner R L, Gharaee-Kermani M, et al. Vitamin A antagonizes decreased cell growth and elevated collagen-degrading matrix metalloproteinases and stimulates collagen accumulation in naturally aged human skin. *J Invest Dermatol* 2000; 114: 480-486.
- 5 Cho S, Lowe L, Hamilton T A, Fisher G J, Voorhees J J, Kang S. Long-term treatment of photoaged human skin with topical retinoic acid improves epidermal cell atypia and thickens the collagen band in papillary dermis. *J Am Acad Dermatol* 2005; 53: 769-774.
- 6 Olsen E A, Katz H I, Levine N, et al. Tretinoin emollient cream: a new therapy for photodamaged skin. *J Am Acad Dermatol* 1992; 26: 215-224.
- 7 Machtinger L A, Kaidbey K, Lim J, et al. Histological effects of tazarotene 0.1% cream vs. vehicle on photodamaged skin: a 6-month, multicentre, double-blind, randomized, vehicle-controlled study in patients with photodamaged facial skin. *Br J Dermatol* 2004; 151: 1245-1252.
- 8 Rittie L, Varani J, Kang S, Voorhees J J, Fisher G J. Retinoid-induced epidermal hyperplasia is mediated by epidermal growth factor receptor activation via specific induction of its ligands heparin-binding EGF and amphiregulin in human skin in vivo. *J Invest Dermatol* 2006; 126: 732-739.
- 9 Eichner R, Gendimenico G J, Kahn M, Mallon J P, Capetola R J, Mezick J A. Effects of long-term retinoic acid treatment on epidermal differentiation in vivo: specific modifications in the programme of terminal differentiation. *Br J Dermatol* 1996; 135: 687-695.
- 10 Hoal E, Wilson E L, Dowdle E B. Variable effects of retinoids on two pigmented human melanoma cell lines. *Cancer Res* 1982; 42: 5191-5195.
- 11 Kang S, Bergfeld W, Gottlieb A B, et al. Long-term efficacy and safety of tretinoin emollient cream 0.05% in the treatment of photodamaged facial skin: a two-year, randomized, placebo-controlled trial. *Am J Clin Dermatol* 2005; 6: 245-253.
- 12 Zouboulis C C, Korge B, Akamatsu H, et al. Effects of 13-cis-retinoic acid, all-trans-retinoic acid, and isotretinoin on the proliferation, lipid synthesis and keratin expression of cultured human sebocytes in vitro. *J Invest Dermatol* 1991; 96: 792-797.
- 13 Griffiths C E, Russman A N, Majumdar G, Slinger R S, Hamilton T A, Voorhees J J. Restoration of collagen formation in photodamaged human skin by tretinoin (retinoic acid). *N Engl J Med* 1993; 329: 530-535.
- 14 Lowe N, Gifford M, Tanghe E, et al. Tazarotene 0.1% cream versus tretinoin 0.05% emollient cream in the treatment of photodamaged facial skin: a multicenter, double-blind, randomized, parallel-group study. *J Cosmet Laser Ther* 2004; 6: 79-85.
- 15 Tagami H, Tadaki T, Obata M, Koyama J. Functional assessment of the stratum corneum under the influence of oral aromatic retinoid (tretinate) in guinea-pigs and humans. Comparison with topical retinoic acid treatment. *Br J Dermatol* 1992; 127: 470-475.
- 16 Varani J, Fligiel H, Zhang J, et al. Separation of retinoid-induced epidermal and dermal thickening from skin irritation. *Archives of dermatological research* 2003; 295: 255-262.
- 17 Wood L C, Elias P M, Calhoun C, Tsai J C, Grunfeld C, Feingold K R. Barrier disruption stimulates interleukin-1 alpha expression and release from a pre-formed pool in murine epidermis. *J Invest Dermatol* 1996; 106: 397-403.
- 18 Blanton R A, Kupper T S, McDougall J K, Dowler S. Regulation of interleukin 1 and its receptor in human keratinocytes. *Proc Natl Acad Sci U S A* 1989; 86: 1273-1277.
- 19 Ya-Xian Z, Suetake T, Tagami H. Number of cell layers of the stratum corneum in normal skin - relationship to the anatomical location on the body, age, sex and physical parameters. *Arch Dermatol Res* 1999; 291: 555-559.
- 20 de Jongh C M, Jakasa I, Verberk M M, Kezic S. Variation in barrier impairment and inflammation of human skin as determined by sodium lauryl sulphate penetration rate. *Br J Dermatol* 2006; 154: 651-657.
- 21 Jacobson E L, Kim H, Kim M, Wondrak G T, Jacobson M K. Developing topical prodrugs for skin cancer prevention. In: Alberts D S, Hess L M, eds. *Fundamentals of Cancer Prevention*. Berlin/Heidelberg: Springer-Verlag, 2005: 139-150.
- 22 Jacobson E L, Kim H, Kim M, et al. A topical lipophilic niacin derivative increases NAD, epidermal differentiation, and barrier function in photodamaged skin. *Exp Dermatol* 2007; 16: 490-499.
- 23 Fitzpatrick T B. The validity and practicality of sun-reactive skin types I through VI. *Arch Dermatol* 1988; 124: 859-871.
- 24 Glogau R G. Aesthetic and anatomic analysis of the aging skin. *Semin Cutan Med Surg* 1996; 15: 134-138.
- 25 Matkowskyj K A, Cox R, Jensen R T, Benya R V. Quantitative immunohistochemistry by measuring cumulative signal strength accurately measures receptor number. *J Histochem Cytochem* 2003; 51: 205-214.
- 26 Fuhr J W, Feingold K R, Elias P M. Transepidermal water loss reflects permeability barrier status: validation in human and rodent in vivo and ex vivo models. *Exp Dermatol* 2006; 15: 483-492.
- 27 Jakasa I, de Jongh C M, Verberk M M, Bos J D, Kezic S. Percutaneous penetration of sodium lauryl sulphate is increased in uninvolved skin of patients with atopic dermatitis compared with control subjects. *Br J Dermatol* 2006; 155: 104-109.
- 28 Gibbs S, Backendorf C, Ponc M. Regulation of keratinocyte proliferation and differentiation by all-trans-retinoic acid, 9-cis-retinoic acid and 1,25-dihydroxy vitamin D3. *Arch Dermatol Res* 1996; 288: 729-738.
- 29 Monteiro-Riviere N A, Bristol D G, Manning T O, Rogers R A, Riviere J E. Interspecies and interregional analysis of the comparative histologic thickness and laser Doppler blood flow measurements at five cutaneous sites in nine species. *J Invest Dermatol* 1990; 95: 582-586.
- 30 Bhawan J, Olsen E, Luffano L, Thorne E G, Schwab B, Gilchrist B A. Histologic evaluation of the long term effects of tretinoin on photodamaged skin. *J Dermatol Sci* 1996; 11: 177-182.
- 31 Weinstein G D, Nigra T P, Pochl P E, et al. Topical tretinoin for treatment of photodamaged skin. A multicenter study. *Arch Dermatol* 1991; 127: 659-665.
- 32 Tur E, Hohl D, Jetten A, Panizzon R, Frenk E. Modification of late epidermal differentiation in photoaged skin treated with topical retinoic acid cream. *Dermatology* 1995; 191: 124-128.
- 33 Elias P M. Stratum corneum defensive functions: an integrated view. *J Invest Dermatol* 2005; 125: 183-200.
- 34 Segre J A. Epidermal barrier formation and recovery in skin disorders. *J Clin Invest* 2006; 116: 1150-1158.
- 35 Wickert R R, Visscher M O. Structure and function of the epidermal barrier. *Am J Infect Control* 2006; 34: S98-S110.
- 36 Levine N, Moon T E, Carimel B, et al. Trial of retinol and isotretinoin in skin cancer prevention: a randomized, double-blind, controlled trial. Southwest Skin Cancer Prevention Study Group. *Cancer Epidemiol Biomarkers Prev* 1997; 6: 957-961.
- 37 Hageman G J, Stierum R H. Niacin, poly(ADP-ribose) polymerase-1 and genomic stability. *Mutat Res* 2001; 475: 45-56.
- 38 Kirkland J B. Niacin and carcinogenesis. *Nutr Cancer* 2003; 46: 110-118.

DOI:10.1111/j.1600-0625.2007.00553.x  
www.blackwellpublishing.com/EXD

Original Article

## A topical lipophilic niacin derivative increases NAD, epidermal differentiation and barrier function in photodamaged skin

Elaine L. Jacobson<sup>1,2,3</sup>, Hyuntae Kim<sup>1,3</sup>, Moonsun Kim<sup>1,3</sup>, Joshua D. Williams<sup>1,3</sup>, Donna L. Coyle<sup>1,3</sup>, W. Russell Coyle<sup>1,3</sup>, Gary Grove<sup>4</sup>, Ronald L. Rizer<sup>5</sup>, M. Suzanne Stratton<sup>2</sup> and Myron K. Jacobson<sup>1,2,3</sup>

<sup>1</sup>Department of Pharmacology and Toxicology, College of Pharmacy, University of Arizona, Tucson, AZ, USA;

<sup>2</sup>Arizona Cancer Center, University of Arizona, Tucson, AZ, USA;

<sup>3</sup>Niadyme Development, Inc., Tucson, AZ, USA;

<sup>4</sup>Skin Study Center, Broomall, PA, USA;

<sup>5</sup>Thomas Stephens & Associates, Inc., Colorado Springs, CO, USA

Correspondence: Elaine L. Jacobson, PhD, Arizona Cancer Center, 1515 N. Campbell Ave, Tucson, AZ 85724, USA, Tel.: +1 520 626 5953, Fax: +1 520 626 8567, e-mail: elaine.jacobson@pharmacy.arizona.edu

Accepted for publication: 2 February 2007

**Abstract** The effects of myristyl nicotinate (MN), a nicotinic acid derivative designed to deliver nicotinic acid to skin without vasodilatation, on subjects with photodamaged skin have been studied. MN increased skin cell nicotinamide adenine dinucleotide (NAD) by 25% ( $P = 0.001$ ) demonstrating effective delivery of nicotinic acid to skin. Relative to placebo, MN treatment of photodamaged facial skin increased stratum corneum thickness by approximately 70% ( $P = 0.0001$ ) and increased epidermal thickness by approximately 20% ( $P = 0.001$ ). In two separate studies, MN treatment increased rates of epidermal renewal by 6% ( $P = 0.003$ ) to 11% ( $P = 0.001$ ) and increased the minimal erythral dose by 8.9 ( $P = 0.07$ ) and 10% ( $P = 0.05$ ) relative to placebo. MN treatment resulted in reductions in the rates of transepidermal water loss (TEWL) of approximately 20% relative

to placebo on cheeks ( $P = 0.012$ ) and arms ( $P = 0.017$ ) of study subjects. Results of a tape stripping challenge before and after MN treatment demonstrated a significant correlation ( $P = 0.03$ ) between increased skin NAD content and resistance to changes in TEWL for MN treated but not placebo subjects. Rates of TEWL changed more rapidly and to a greater extent in atopic subjects compared with normal subjects. The results indicate that MN enhances epidermal differentiation and barrier function in skin, suggesting that this method of nicotinic acid delivery may prove useful in limiting progression of actinic skin damage and possibly in treating other conditions involving skin barrier impairment.

**Key words:** atopic skin – epidermal differentiation – niacin/NAD – photodamage – skin barrier function

Please cite this paper as: A topical lipophilic niacin derivative increases NAD, epidermal differentiation and barrier function in photodamaged skin. *Experimental Dermatology* 2007; 16: 490–499.

### Introduction

Chronic solar exposure to skin (photoaging) is associated with multiple alterations in structure and function (1–3). These changes include rearrangement of collagen and elastic fibres in the extracellular matrix of the dermis, irregularities in melanocyte and keratinocyte morphology in the epidermis (4), decreases in stratum corneum thickness, flattening of the dermal–epidermal junction (1, 5), and impairment of skin barrier function (6–8). Early stage pho-

todamage can result in skin hyperpigmentation, loss of skin smoothness and wrinkling while later stage photodamage can result in development of actinic (solar) keratosis, a hyperproliferative lesion thought to be a continuum with non-melanoma skin cancer (9), and to melanoma skin cancer (10). While use of sunscreens represents a front line strategy for limiting actinic damage, their efficacy is still limited by inadequate use, incomplete spectral protection and potential dermal toxicities (11). Therefore, other approaches to limit the progression of skin damage could complement the use of sunscreens (12, 13).

Preclinical studies suggest that nicotinic acid may provide benefit to photodamaged skin by mechanisms that involve both nutrient and drug effects of this agent. Regarding nutrient effects, nicotinic acid and the other B3

**Abbreviations:** H&E, haematoxylin–eosin; MED, minimal erythral dose; MN, myristyl nicotinate; PARP, poly(ADP-ribose) polymerase; ROS, reactive oxygen species; TEWL, transepidermal water loss; UV, ultraviolet light.

vitamer, niacinamide, can serve as precursors to nicotinamide adenine dinucleotide (NAD), albeit by different metabolic pathways (14). The oxidized form of NAD is consumed as a substrate in poly(ADP-ribose) polymerase-1/2 (PARP-1/2) catalyzed reactions that promote genomic stability following DNA damage such as that caused by ultraviolet (UV) light (15, 16). UV exposure can lead to depletion of NAD in skin cells and intact skin, resulting in insufficient NAD available for PARP-1/2 activity (17, 18). Supplementation with nicotinic acid at levels that increase skin NAD content inhibits UV-induced carcinogenesis and photoimmune suppression in an animal model (19). NAD functions as a hydride ion acceptor and donor in biological redox reactions central to energy metabolism, and as a large organ with a high rate of turnover, the epidermal compartment of skin has a high-energy requirement (20). NAD is the precursor for NADPH (21), which is essential for synthesis of many lipids including ceramides important in skin barrier function (22), and serves as the source of reducing equivalents for cellular antioxidants needed to counter UV induced reactive oxygen species (23). Regarding drug effects, a G-protein coupled nicotinic acid receptor (24, 25), present in skin (26), is likely involved in release of the cytokine leptin (27). In skin, leptin-mediated signalling pathways show protective effects that include enhancing epidermal differentiation (28–30) and wound healing (31, 32).

We report here clinical evaluations of myristyl nicotinate (MN), a lipophilic derivative of nicotinic acid developed to deliver nicotinic acid to skin following topical application (33). Our results indicate that this approach to deliver nicotinic acid to skin offers a new tool for management of skin photodamage. Additionally, the ability to enhance skin barrier function suggests that MN may benefit other dermatology conditions that involve barrier impairment.

## Materials and methods

### Overview of clinical studies

Data obtained from three clinical studies are described in this communication. All studies were conducted in accordance with applicable Good Clinical Practice regulations and guidelines and Institutional Review Board (IRB) regulations. All subjects were required to read and sign an IRB-approved Informed Consent Form. Sample size was determined empirically. Each subject who qualified for enrollment was assigned a subject number that was used on all subject documentation. These unique numbers were assigned in ascending order using a computer-generated randomization schedule developed by the contract research organization (CRO) conducting each trial. Numbers were concealed until after the intervention was assigned. Each CRO enrolled and assigned subjects to groups. All partici-

pants, those administering the interventions, and those assessing the outcomes were blinded to group assignment. Analyses were not by 'intention-to-treat' but only on those who completed the study. Two occurrences of allergic or adverse reactions to the formulations were noted in the 96 subjects who enrolled in these studies. Both of these were classified by clinical evaluators as a mild rash appearing within 24 h of the first application, possibly related to study product, which resolved in three and 9 days, respectively, and both subjects continued in the study. Analysis was restricted to subjects with complete datasets, thus the *n* for some measures differs slightly because of missing data points.

### Study 1

Study 1 was an 18-week assessment (12/06/00–04/12/01) conducted by the Skin Study Center, Broomall, Pennsylvania, USA. It involved 16 healthy adult females between 35 and 45 years old with Fitzpatrick Skin Types I–IV. Subjects applied a simple, oil in water lotion containing 1% MN to one total forearm (complete coverage from wrist to elbow) and placebo lotion (myristyl myristate in the placebo replaced MN contained in the active) to the other total forearm in a double-blinded protocol. All applications were 0.5 ml once per day per site (1 mg/cm<sup>2</sup>). Individuals were ineligible if, in the opinion of the investigator, they had known allergies or sensitivities to products; exhibited any skin disorders on the test areas of the face, upper inner arms or volar forearms; had known medical conditions, such as diabetes, that could affect wound healing or; were using medications which might have influenced the study (e.g. prescription strength steroids, anti-inflammatory drugs or topical medications). Subjects were ineligible if they were pregnant or nursing.

### Study 2

Study 2 was a 12-week randomized, double-blind, placebo-controlled evaluation of the effects of a 5% MN formulation on human skin NAD and photoaging parameters. It was conducted (12/12/02–04/07/03) by Thomas J. Stephens & Associates, Colorado Springs, CO, USA. Female subjects ages 35–60, with a Fitzpatrick Skin Classification score of I–IV, moderate to advanced photodamaged skin defined by a Modified Glogau Classification of I–III (34), and significant dyschromia on the face determined by a score of  $\geq 4$  on the 10-cm visual acuity scale, participated in the study. The mean age was  $48.2 \pm 6.8$  and 85% were Caucasian and 15% Hispanic. Subjects were randomly assigned to one of two groups of 30 (active or placebo where the formulations were identical except that myristyl myristate in the placebo replaced MN contained in the active). Test materials were applied to face and total forearms of subjects each morning and evening, 0.5 ml to the face (1 mg/cm<sup>2</sup>) and 1 ml to

each arm ( $2 \text{ mg/cm}^2$ ). A separate group of atopic subjects, qualified for a substudy by having confirmed atopic criteria based on a questionnaire validated by the CRO, participated only in TEWL measurements on the volar forearms. Subjects were required to be willing to avoid direct sun exposure on the forearms and the use of tanning beds for the duration of the study. Individuals were ineligible if, in the opinion of the investigator, they had known allergies or sensitivities to products that may have influenced the study; exhibited any skin disorders on the test areas of the face, upper inner arms or volar forearms; had known medical conditions, such as diabetes, that could affect wound healing or; were using medications which might have influenced the study (e.g. prescription strength steroids, anti-inflammatory drugs, or topical medications). Subjects were ineligible if they were pregnant or nursing. Other exclusion criteria included; hypertension or uncontrolled metabolic disease, such as thyroid disease; known sensitivity to alpha- and beta-hydroxyacids or lactic acid; use of products containing hydroxy acids, or retinoids within 1 month of enrollment; participation in dansyl chloride patch-type studies within 24 months of enrollment; and/or concurrent participation in clinical studies involving the arms.

#### Study 3

Study 3 involved an 8-week double-blinded, placebo-controlled study to evaluate biophysical effects of formulations containing niacin derivatives on skin. The subjects were females in generally good health with Modified Glogau Classification, I-II; Fitzpatrick Classification, I-III, between the age of 30 and 60 years with a mean age of  $44.1 \pm 7.3$  where 92% were Caucasian, 6% were Hispanic and 2% were Hispanic/Caucasian. The study was conducted (06/14/05–10/08/05) by Thomas J. Stephens & Associates, Colorado Springs, CO, USA. Twenty subjects were examined at baseline and weeks 2, 4, and 8 during use of a 5% MN formulation applied to one total forearm and a placebo formulation (formulations were identical to those in Study 2) applied to the other total forearm (arm assignment based on a predetermined randomization provided by the test facility) ( $2 \text{ mg/cm}^2$ ). Inclusion and exclusion criteria and doses were the same as described in Study 2.

#### Skin biopsy analyses

A board-certified dermatologist collected a 4-mm punch biopsy from the dorsal forearm of 20 randomly selected subjects (nine in placebo and 11 in the MN group) at baseline and after 12 weeks of treatment in Study 2. Samples were stored at  $-80^\circ\text{C}$  and NAD was measured as reported previously (35). A 2-mm periorbital punch biopsy was taken by a board-certified dermatologist at baseline (right side) and 12 weeks (left side) on randomly selected subjects in Study 2. The biopsies were formalin-fixed, embedded in paraffin,

cut into  $5\text{-}\mu\text{m}$  cross-sections, mounted on slides, and stained with haematoxylin-eosin (H&E). Images were taken of H&E-stained cross-sections with a Nikon Eclipse TE300 microscope using a  $10\times$  by 0.45 Apochromat objective and a Coolsnap Photometrics digital CCD camera. IMAGEJ software (NIH) was used to analyze the images. Suprapapillary epidermal thickness (as measured from the top of the dermal papilla to the top of the granular layer) and stratum corneum thickness (as measured from the top of the granular layer to the top of the stratum corneum) were measured at five different sites and the average was calculated.

#### Stratum corneum turnover determination

Application of test formulations to the right and left upper inner arms began following baseline assessments. A 5% dansyl chloride suspension in white petrolatum was applied under occlusion on each site after 4 weeks of treatment. Six hours later, a second application was made and the dressings renewed. After 24 h, the occlusive dressings were removed and the test sites were washed. After drying, each site was examined under a Wood's lamp and the acceptability of staining determined. Application of test formulations resumed on the evening the dye patches were removed. The degree of residual staining from the dansyl chloride was assessed three times per week by a clinician in blinded fashion for 4 weeks or until all sites were no longer fluorescent under Wood's lamp illumination.

#### Minimal erythema dose (MED) determination

Minimal erythema dose is defined here as the time of light exposure producing a minimally perceptible erythema reaction discernible 16–24 h after irradiation with a single port solar simulator (Solar Light Co., Philadelphia, PA, USA) equipped with a 150-watt xenon arc lamp (Model 16S, Solar UV Simulator, Solar Light Co.). A spectral output similar to that of the natural solar spectrum (emissions of UVA+B, 290–400 nm) was obtained by using a combination of the UG-5 or UG-11 and WG-320 filters (Schott Glass Technologies, Elmsford, NY, USA). The output of the solar simulator was monitored with a 3D-600 m (Solar Light Co.), and calibrations occurred just prior to use. After 18 weeks of test formulation use in Study 1 and 12 weeks of use in Study 2, MED was determined by exposing six squares of skin on the volar forearms to varying doses of UV. Each irradiated site received 25% more exposure than the previous site, starting at 0.64 of the estimated MED for that skin type (0.64, 0.8, 1.0, 1.25, 1.56 and 1.95 times the estimated MED) and 16–24 h later, the MED was determined by the site that exhibited the least perceptible erythema.

#### Transepidermal water loss (TEWL) measurement

Subjects were equilibrated to ambient conditions for at least 20 min and maintained between  $66$  and  $72^\circ\text{F}$  and rel-

ative humidity of 15–55%. A Dermalab instrument was used to measure TEWL at two points on the right volar forearm and left outer lower cheek, averaged over a 1-min measurement period.

### Tape-stripping challenge

The right anterior volar forearm was compromised by applying acetone to the forearm for 30 s, then applying and removing 10 strips of 1" Blenderm tape in a cross-wise pattern. TEWL measurements were made before and after tape stripping as described above.

## Results

### A topical MN formulation elevates NAD, a biomarker of niacin delivery, in human skin

Skin NAD content was used as a biomarker of nicotinic acid delivery following topical application of formulations containing MN. NAD provides a biomarker for all phases of delivery including partitioning from stratum corneum to living layers of the epidermis, conversion to nicotinic acid by skin esterases, and cellular uptake and bioconversion to NAD. Twenty subjects completed a phase of Study 2 where 4-mm punch biopsies of the forearm were obtained at baseline and after 12 weeks use of test formulations. These subjects were randomly assigned to placebo ( $n = 9$ ) and 5% MN groups ( $n = 11$ ). Biopsies were processed and analyzed for skin NAD content, expressed as pmol NAD per microgram protein (Fig. 1). At baseline, the MN group had slightly higher but not statistically significantly different NAD relative to the placebo group. After 12 weeks, no significant increase in NAD was observed in the placebo group, while the MN group increased skin NAD to an average of 125% of baseline ( $P = 0.016$ ). The difference in NAD between placebo and MN groups at 12 weeks was highly statistically significant ( $P = 0.008$ ). These data demonstrate that MN delivered nicotinic acid to skin in a manner that allowed conversion to nicotinic acid and cellular uptake and bioconversion to NAD.

### MN promotes epidermal differentiation in photodamaged skin without increasing photosensitivity

The effect of MN on epidermal histology in photodamaged skin was evaluated by comparison of cheek biopsy samples from placebo and MN-treated subjects from Study 2 at baseline and after 12 weeks of application. Increases of stratum corneum thickness are associated with increased epidermal differentiation and impaired differentiation resulting in decreased stratum corneum thickness occurs in atopic dermatitis (36). Visible increases in the stratum corneum thickness and in epidermal thickness as a result of increased cellularity were observed for subjects receiving MN. An example is shown in Fig. 2, where (a) shows a pla-

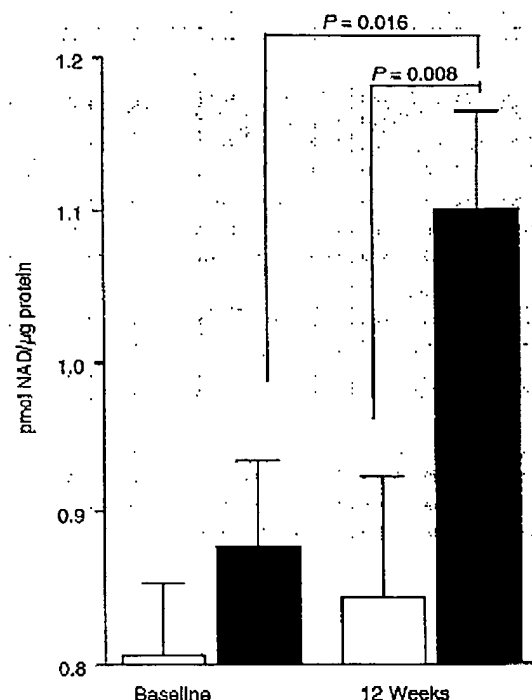
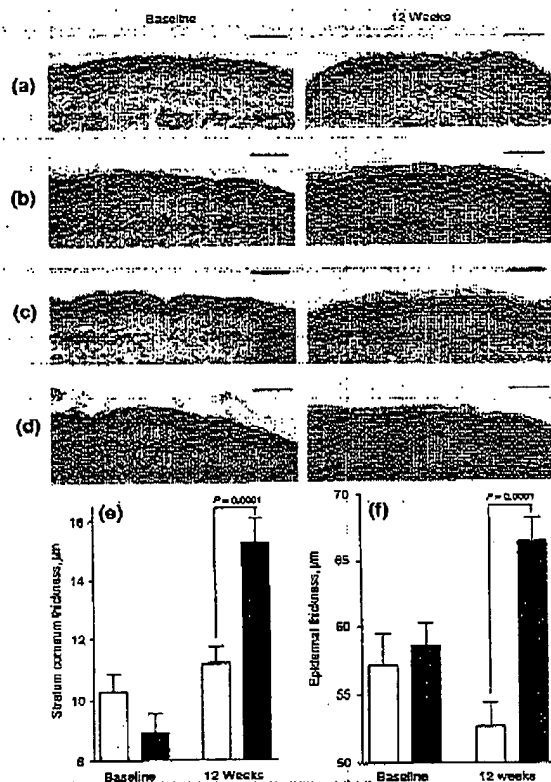


Figure 1. Effects of myristyl nicotinate on skin nicotinamide adenine dinucleotide (NAD) content. Skin punch biopsies obtained in Study 2 at baseline and 12-weeks post-treatment from the forearm were analyzed for NAD content, which is expressed relative to protein. Solid bars represent subjects randomized to myristyl nicotinate treatment ( $n = 11$ ) and open bars represent subjects in the placebo group ( $n = 9$ ). The values shown are group means  $\pm$  SEM and the  $P$ -values were derived from a paired, two-tailed Student's  $t$ -test for within group comparisons and unpaired analyses for between group comparisons.

cebo subject and (b–d) show subjects that applied formulations containing 5% MN. Data summarizing changes in stratum corneum and epidermal thickness for all subjects (Fig. 2e and f) demonstrated a highly significant difference from baseline to 12 weeks between MN and placebo-treated groups for both parameters ( $P = 0.0001$ , both parameters,  $n = 31$  for MN and 27 for placebo). The mean increase in stratum corneum thickness in the MN group was approximately 70% ( $6.35 \mu\text{m}$ ) while the placebo group increased only slightly ( $0.89 \mu\text{m}$ ). Epidermal thickness in the placebo group decreased by an average of  $4.47 \mu\text{m}$  (approximately 8%) while the MN group increased in thickness by  $7.75 \mu\text{m}$  (approximately 13%), a differential effect of approximately 20%.

Two studies examined the effect of MN on the rate of stratum corneum turnover following staining with dansyl chloride as a surrogate marker of epidermal renewal. Study 1 involved treatment with a 1% MN formulation once per day and Study 2 involved treatment with a 5% formulation

Jacobson et al.



**Figure 2.** Effects of myristyl nicotinate (MN) on stratum corneum and epidermal thickness. Representative histological analyses of placebo and MN-treated skin biopsies: H&E slides were prepared from peri-ocular skin punch biopsies from subjects in Study 2. H&E from a representative placebo subject (a) at baseline and 12 weeks of treatment and from three representative subjects treated with MN (b–d) at baseline and 12 weeks of treatment. The bar in the upper right corner represents 100 microns. In (e), the mean stratum corneum thicknesses  $\pm$  SEM for placebo ( $n = 27$ ) and MN ( $n = 31$ ) groups are shown at baseline and after 12 weeks of treatment. In (f), the mean epidermal thicknesses  $\pm$  SEM for placebo ( $n = 27$ ) and MN ( $n = 31$ ) groups are shown at baseline and after 12 weeks. The  $P$ -values shown are derived from an unpaired, two-tailed  $t$ -test of change from baseline to 12 weeks between groups.

twice per day. In both studies, formulations were applied for 4 weeks prior to assessment of turnover time by evaluation of dansyl fluorescence extinction on alternate days for 4 weeks starting at day 6 after application to determine the  $t_{1/2}$  for turnover. The presence of MN increased the rate of dansyl turnover relative to placebo by approximately 11.3% in Study 1 ( $P = 0.001$ ) and 6.3% ( $P = 0.003$ ) in Study 2 (Table 1), demonstrating enhanced epidermal renewal by MN.

As topical formulations that stimulate epidermal turnover have been shown to result in the occurrence of photosensitivity (37), the effect of MN on the minimum time of UV exposure to elicit erythema (MED) was measured in

both Study 1 and 2. The results (Table 1) show that MN formulations increased the MED by approximately 9–10% compared with placebo. The results from Study 1 approached but did not reach statistical significance ( $P = 0.07$ ,  $n = 16$ ), while the data from Study 2 did reach statistical significance at  $P = 0.05$  ( $n = 24$ ). These studies demonstrate that use of MN does not result in development of photosensitivity but rather that its use can provide mild photoprotection.

### MN enhances skin barrier function

The importance of the skin barrier in normal homeostasis and its impairment in a number of dermatology conditions are the areas of considerable current research in dermatology (38, 39). Stratum corneum thickness has been identified as an important determinant of skin barrier function (40) and the ability of MN to increase stratum corneum thickness in photodamaged skin (Fig. 2) along with providing photoprotection (Table 1) suggested that this compound may be enhancing skin barrier function. Accordingly, effects of MN on skin barrier function were examined by using three different approaches.

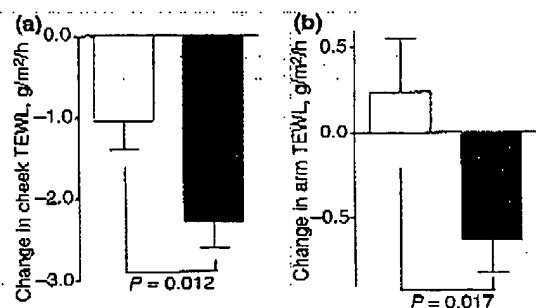
Changes in the rates of TEWL have long been used as a marker of changes in skin barrier function (38, 40–42). Accordingly, changes in rates of TEWL on both the face and arms of subjects in Study 2 were determined at baseline and after 12 weeks for both placebo and MN formulations (Fig. 3). Skin cream moisturizers in general have been shown to increase barrier function (43, 44) and the placebo formulation in this study applied to the face of study subjects resulted in decreased rates of TEWL of 1.0 g/m<sup>2</sup>/h, which corresponded to a decrease of approximately 11%. However, formulations containing 5% MN decreased water loss by approximately 2.5 g/m<sup>2</sup>/h or 250% relative to the placebo formulation ( $P = 0.012$ ,  $n = 29$  for placebo and 31 for MN groups). The rates of TEWL on the arms of placebo subjects increased over the course of the study while the MN group decreased, but again a highly statistically significant differential effect between placebo and MN groups ( $P = 0.017$ ,  $n = 28$  for placebo and 30 for MN groups) was observed. The increase in rates of TEWL on the arms of placebo subjects likely reflects seasonal changes in skin barrier function, which occurred between December and April in the study shown.

A second approach to examine effects of MN on skin barrier function involved a study of the effects of tape stripping on changes in the rates of TEWL. In this study, the rates of TEWL were determined both before and following a standard regimen of tape stripping at both baseline and after 12 weeks. Prior studies have shown that a strengthened skin barrier is more resistant to increases in rates of TEWL following a standard regimen of tape stripping (45). The changes in TEWL values ( $\Delta$ TEWL) at 12 weeks minus the

**Table 1.** Effect of myristyl nicotinate (MN) on epidermal renewal and MED

MN	n	Placebo	n	MN-dependent change	P-value
Dansyl turnover time, $t_{1/2}$ (days)					
Study 1 (1%) $17.2 \pm 0.44$	16	$19.4 \pm 0.41$	16	11.3%	0.001
Study 2 (5%) $13.4 \pm 0.185$	34	$14.3 \pm 0.226$	34	6.44%	0.003
MED (s)					
Study 1 (1%) $29.4 \pm 2.6$	16	$27.0 \pm 1.8$	16	8.9%	0.07
Study 2 (5%) $48.7 \pm 1.8$	24	$44.3 \pm 0.41$	24	10.0%	0.05

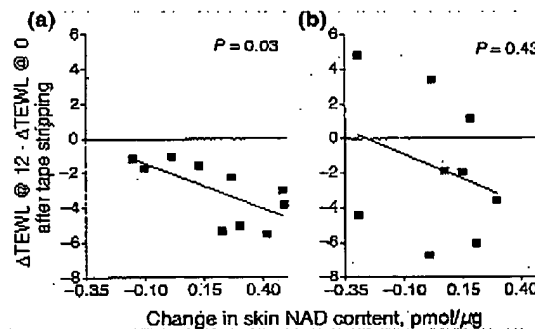
The  $t_{1/2}$  for epidermal renewal was measured as described in Materials and methods after 4-week use of formulations. MED was determined as described in Materials and methods.



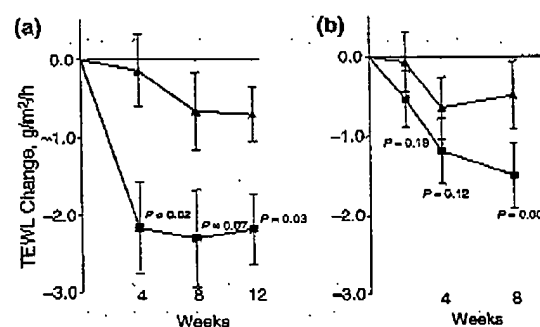
**Figure 3.** Effects of a 5% myristyl nicotinate (MN) formulation on skin barrier function assessed by changes in rates of transepidermal water loss (TEWL) on the face and volar forearms of subjects in Study 2. Panel (a) shows the change in TEWL rates on the cheek from baseline to 12 weeks while (b) shows the same measurements on the volar forearms. Open bars represent the placebo group and the black bars represent the MN-treated group. The values are mean  $\pm$  SEM where,  $n = 29$  and  $28$  for the placebo group in A and B, respectively, and  $31$  and  $30$  for the MN-treated groups, respectively. The  $P$ -values are derived from unpaired, two-tailed Student's  $t$ -tests.

$\Delta$ TEWL values at baseline ( $\Delta$ TEWL<sub>12</sub> -  $\Delta$ TEWL<sub>0</sub>) were then plotted as a function of the change in skin NAD, which serves as a biomarker of delivery by MN (Fig. 4). In this plot, a value of zero represents no change in barrier function between baseline and 12 weeks, negative values indicate an improvement in barrier function and positive values indicate a decrease in barrier function. The results for the MN-treated group (panel a) show that all subjects improved in barrier function ( $\Delta$ TEWL<sub>12</sub> -  $\Delta$ TEWL<sub>0</sub> < 0). Additionally, a strong correlation between the change in skin NAD and  $\Delta$ TEWL<sub>12</sub> -  $\Delta$ TEWL<sub>0</sub> values (Pearson  $r = -0.68$ ,  $P = 0.03$ ,  $n = 10$ ) was observed for the MN group. Correlation analysis (panel b) did not show a significant correlation in the placebo group between change in NAD and  $\Delta$ TEWL<sub>12</sub> -  $\Delta$ TEWL<sub>0</sub> (Pearson  $r = -0.13$ ,  $P = 0.43$ ,  $n = 9$ ).

Studies have shown that rates of TEWL are greater in subjects with atopic skin (38, 46, 47). Accordingly, changes in the rates of TEWL in a group of atopic subjects were compared with a normal group (Fig. 5). While the effects of the placebo formulation were similar in the two populations,



**Figure 4.** Correlation analysis of change in skin nicotinamide adenine dinucleotide (NAD) content and changes in TEWL before and after a tape stripping challenge from Study 2. The changes in transepidermal water loss (TEWL) values ( $\Delta$ TEWL) at baseline minus the  $\Delta$ TEWL values at 12 weeks ( $\Delta$ TEWL<sub>12</sub> -  $\Delta$ TEWL<sub>0</sub>) are plotted as a function of the change in skin NAD, which serves as a biomarker of delivery by MN. Panel (a) shows the data for MN-treated subjects and (b) shows the data for the placebo group where  $n = 10$  and  $9$  subjects, respectively. The Pearson  $r$ -value for (a) =  $-0.68$  and for (b) =  $-0.13$ .  $P$ -values were derived from the same analysis as for the Pearson  $r$ -value.



**Figure 5.** Comparison of effects of myristyl nicotinate (MN) on changes in the rates of transepidermal water loss (TEWL) in atopic and non-atopic subjects. The time course of TEWL changes on the forearms is shown in (a) for atopic subjects from Study 2 treated twice daily with placebo (triangles,  $n = 6$ ) and 5% MN (squares,  $n = 7$ ) and shown in (b) for non-atopic subjects in Study 3 treated twice daily with placebo (triangles,  $n = 24$ ) and 5% MN (squares,  $n = 24$ ). The  $P$ -values are derived from a paired two-tailed Student's  $t$ -test.



rates of TEWL decreased more rapidly and to a greater extent in atopic subjects than in non-atopic subjects. At 4 weeks, the first measurement made in atopic individuals, a highly significant decrease in TEWL was observed in treated subjects relative to the placebo group ( $P = 0.02$ ,  $n = 7$  and 6, respectively.) while statistically significant differences were not observed until 8 weeks in the normal subjects.

## Discussion

A number of preclinical studies indicating that nicotinic acid could benefit photodamaged skin by several possible mechanisms provided the basis for the clinical studies described here. A major obstacle to the delivery of therapeutic amounts of nicotinic acid to any tissue is its ability to cause a peripheral vasodilatation that leads to a skin flushing response, an effect that is not harmful but is intensely disliked by most patients (48, 49). Delivery of nutrients taken orally to the epidermal compartment of skin is inherently inefficient and, combined with skin flushing side effects, oral delivery of nicotinic acid to skin with high patient compliance seems unlikely. Likewise, topical application of formulations containing nicotinic acid per se at levels above 0.05% is not feasible because they elicit intense vasodilatation at the site of application (33). The feasibility of topical delivery of nicotinic acid without causing vasodilatation was established by demonstrating in animal models that delivery of nicotinic acid by topical application of long chain ester derivatives such as MN can achieve delivery of relatively large amounts of nicotinic acid without skin flushing. The rate of diffusion of the ester derivative from the stratum corneum decreases as its lipophilicity increases, allowing the rate of delivery to be reduced such that the levels of free nicotinic acid released by skin esterases remain below the threshold for vasodilatation (33). Recent studies indicate that the skin flushing response is mediated by activation of receptors on epidermal Langerhans cells (50) or macrophages at concentrations above 0.1 mM (51). The topical application of formulations containing up to 5% MN did not induce skin flushing in the subjects involved in these trials and even higher concentrations do not cause this effect (data not shown). Using skin NAD as a biomarker of nicotinic acid delivery, MN was shown to deliver nicotinic acid to skin (Fig. 1) in human subjects effectively. As the method assesses intracellular NAD, the increases are likely derived primarily from the epidermis where the density of cells is much greater than in the dermis. However, determining which skin layer is most affected is difficult as the method for measurement requires intact cells where NADase activity has not been activated by disruption of the tissue.

Chronic sun exposure to skin often leads to thinning of the stratum corneum, sometimes referred to as compaction

(1). Our studies (Fig. 2) show that MN treatment leads to an increase in stratum corneum thickness in photodamaged facial skin that averaged approximately 70% in the population studied. In the study reported here, changes in stratum corneum thickness have been determined in paraffin-embedded samples subjected to the same conditions of preparation as the placebo samples. While prior studies have shown that the thickness of the stratum corneum is less in paraffin sections compared with frozen sections (52), paraffin sections have been used to assess stratum corneum morphology and changes in thickness in numerous studies (53–57). The relative changes in stratum corneum thickness observed in this study strongly suggest that MN promotes epidermal differentiation in photodamaged skin. This conclusion also is supported by MN-stimulated increases in epidermal cellularity and thickness (Fig. 2) and rates of epidermal renewal (Table 1). The effect of MN on biomarkers of epidermal differentiation will be reported elsewhere (E. L. Jacobson et al. in preparation). The ability of MN to stimulate epidermal renewal without causing photosensitivity (Table 1) is in contrast to the effects of other agents that stimulate epidermal differentiation. An example is retinoic acid treatment where stimulation of epidermal renewal is accompanied by increased photosensitivity (37).

Recent studies have identified stratum corneum thickness as a key component of skin barrier function (40, 42), thus the ability of MN to increase stratum corneum thickness (Fig. 2) provided an indication that it may improve skin barrier function. TEWL measurements have been widely used in the non-invasive assessment of stratum corneum function and the utility of changes in TEWL as a reflection of changes in permeability barrier status has been validated (47). The absolute values of TEWL vary with the site of measurement, age of the individual and the presence of atopic skin (58, 59). In the present study, changes in rates of TEWL over time were assessed as a measure of change in barrier function. Consistent with other studies (43, 44), our results show that daily use of a skin moisturizer can improve the barrier function of photodamaged skin as evidenced by decreases in TEWL observed with placebo formulations (Fig. 3a). However, marked additional decreases in TEWL observed in subjects using MN indicate that this agent provides additional skin barrier enhancement, which together with the observed increases in stratum corneum thickness leads to the conclusion that MN improves barrier function in photodamaged skin. Additionally, the studies reported here indicate that MN also improves barrier function in relatively sun-protected skin as rates of TEWL were reduced on volar forearms of normal (Figs 3b–5b) and atopic subjects (Fig. 5a). The ability of skin to resist increases in rates of TEWL following a standardized regimen has been used in evaluation of changes of skin barrier function

(45) and this method of evaluation also supports the conclusion that MN improves barrier function (Fig. 4).

While our studies cannot distinguish between nutrient and drug effects of nicotinic acid in stimulation of epidermal differentiation and increased barrier function, prior studies suggest that both mechanisms likely are involved. DNA damage is central to skin photodamage and recent advances in the understanding of the role of niacin-dependent PARPs in DNA repair mechanisms (26) and the requirement for relatively high cellular concentrations of NAD for optimal PARP activity (16, 60) support a possible role of niacin as a protective skin micronutrient. Niacin deficiencies can mimic radiation- and chemical-induced DNA damage in terms of increased single- and double-strand breaks and/or oxidative lesions (61–63). Conversely, enhancing NAD in skin is associated with inhibition of UV-induced skin cancer and immunosuppression (19). Niacin-derived NAD has long been known to be essential in energy production and the constant renewal of the epidermis makes this compartment of skin vulnerable to micronutrient losses and UV exposure can cause further depletion (18). Niacin-derived NADPH is essential in the formation of several lipid components of the stratum corneum that are important determinants of epidermal barrier function (64). Thus, it seems reasonable to postulate that improved niacin status reflected in increased NAD and NADP content would benefit epidermal differentiation and maintenance of the epidermal barrier. Finally, the presence of a nicotinic acid receptor present in skin (26) and the stimulation of leptin release (27) support a drug effect of nicotinic acid as leptin has been shown to exert protective effects that include stimulation of epidermal differentiation, wound healing, and immune function (28–30, 65–67).

It is interesting that formulations containing the other B3 vitamin, nicotinamide, improve rosacea (68) and acne (69) and formulations containing 1-methylnicotinamide improve rosacea (70). Nicotinamide also inhibits melanosome transfer in hyperpigmentation (71) and anti-aging effects of formulations containing nicotinamide have been reported (72). The effects of nicotinamide likely involve mechanisms distinct from MN as topical application of formulations containing nicotinamide did not affect skin NAD content in a similar clinical trial where subjects applied a nicotinamide (4%) containing formulation for 8 weeks (E. L. Jacobson et al., manuscript in preparation). Further, nicotinamide does not bind to the nicotinic acid receptor (25).

The ability of MN to stimulate epidermal differentiation and barrier function in skin suggests that this agent may be useful in limiting the effects of skin photodamage either alone or in combination with other agents. The development of abnormal cell populations that can lead to actinic keratosis and non-melanoma skin cancers involves progressive cellular dedifferentiation. Agents that promote differen-

tiation may limit progression of abnormal cell populations in these clinical conditions. Any prevention strategy requires long-term compliance with therapy, which in turn requires a strong record of safety and tolerability in addition to efficacy. The use of nicotinic acid for modification of blood lipids has established a strong record of safety for this agent (49) and the ability of MN to deliver therapeutic amounts of nicotinic acid to skin without the skin flushing side effects provides tolerability long lacking for this agent. Finally, the ability of MN to stimulate barrier function may benefit other conditions such as atopic dermatitis where skin barrier function is compromised (39, 73).

### Acknowledgements

These studies were funded in part by NIH grant R44 CA-90085 and Niadyne, Inc. Collection, management, analysis and interpretation of the data were done by T. J. Stephens & Associates, Inc. or the Skin Study Center except for measurement of NAD in skin, correlation analysis of NAD and TEWL, and image analysis of biopsies that were done by University of Arizona investigators. Histological processing of biopsies was done by the Cellular Imaging Facility Core of the Southwest Environmental Health Sciences Center (NIH grant ES06694) at the University of Arizona. We thank Tad Dewald for technical assistance with biochemical assays. MKJ and ELJ are principals in Niadyne, Inc., whose sponsored research is managed in accordance with the University of Arizona conflict-of-interest policies.

### References

- 1 Gilchrist B A. A review of skin ageing and its medical therapy. *Br J Dermatol* 1996; 135: 867–875.
- 2 El-Domyati M, Attia S, Saleh F et al. Intrinsic aging vs. photoaging: a comparative histopathological, immunohistochemical, and ultrastructural study of skin. *Exp Dermatol* 2002; 11: 398–405.
- 3 Rabe J H, Mamelak A J, McElguinn P J, Morison W L, Sauder D N. Photoaging: mechanisms and repair. *J Am Acad Dermatol* 2006; 55: 1–19.
- 4 Scharffetter-Kochanek K, Brenneisen P, Wenk J et al. Photoaging of the skin from phenotype to mechanisms. *Exp Gerontol* 2000; 35: 307–316.
- 5 Benedetto A V. The environment and skin aging. *Clin Dermatol* 1998; 16: 129–139.
- 6 Robert M, Bissonauth V, Ross G, Rouabhia M. Harmful effects of UVA on the structure and barrier function of engineered human cutaneous tissues. *Int J Radiat Biol* 1999; 75: 317–326.
- 7 Meguro S, Aral Y, Masukawa K, Uie K, Tokimitsu I. Stratum corneum lipid abnormalities in UVB-irradiated skin. *Photochem Photobiol* 1999; 69: 317–321.
- 8 van den Akker J T, Holroyd J A, Vernon D I, Sterenborg H J, Brown S B. Chronic UVB exposure enhances in vitro percutaneous penetration of 5-aminolevulinic acid in hairless mouse skin. *Lasers Surg Med* 2004; 34: 141–145.
- 9 Ackerman A B, Mones J M. Solar (actinic) keratosis is squamous cell carcinoma. *Br J Dermatol* 2006; 155: 9–22.

10. Berwick M. Pathways to the development of melanoma: a complex issue. *J Invest Dermatol* 2006; **126**: 1932-1933.
11. Damiani E, Rosati L, Castagna R, Carloni P, Greci L. Changes in ultraviolet absorbance and hence in protective efficacy against lipid peroxidation of organic sunscreens after UVA irradiation. *J Photochem Photobiol B* 2006; **82**: 204-213.
12. Guercio-Hauer C, MacFarlane D F, Deleo V A. Photodamage, photoaging and photoprotection of the skin. *Am Fam Physician* 1994; **50**: 327-324.
13. Trautinger F. Mechanisms of photodamage of the skin and its functional consequences for skin ageing. *Clin Exp Dermatol* 2001; **26**: 573-577.
14. Jacobson E L, Lange R A, Jacobson M K. Pyridine nucleotide synthesis in 3T3 cells. *J Cell Physiol* 1979; **99**: 417-425.
15. Hageman G J, Stierum R H. Niacin, poly(ADP-ribose) polymerase-1 and genomic stability. *Mutat Res* 2001; **475**: 45-56.
16. Jacobson E L, Antol K M, Juarez-Salinas H, Jacobson M K. Poly(ADP-ribose) metabolism in ultraviolet irradiated human fibroblasts. *J Biol Chem* 1983; **258**: 103-107.
17. Jacobson E L, Ramasinghani S, Shieh W M. DNA Damage Response Pathways in Human Mammary Epithelial Cells with Sub-Optimal Niacin Status. *Proc Am Assoc Cancer Res* 1999; **40**: 4297.
18. Balard B, Giacomoni P U. Nicotinamide adenosine dinucleotide level in dimethyl sulfate-treated or UV-irradiated mouse epidermis. *Mutat Res* 1989; **219**: 71-79.
19. Gensler H L, Williams T, Huang A C, Jacobson E L. Oral niacin prevents photocarcinogenesis and photoimmunosuppression in mice. *Nutr Cancer* 1999; **34**: 36-41.
20. Jacobson E L, Giacomoni P U, Roberts M J, Wondrak G T, Jacobson M K. Optimizing the energy status of skin cells during solar radiation. *J Photochem Photobiol B* 2001; **63**: 141-147.
21. Kirkland J B. Niacin and carcinogenesis. *Nutr Cancer* 2003; **46**: 110-118.
22. Elias P M, Choi E H. Interactions among stratum corneum defensive functions. *Exp Dermatol* 2005; **14**: 719-726.
23. Wondrak G T, Jacobson M K, Jacobson E L. Endogenous UVA-photosensitizers: mediators of skin photodamage and novel targets for skin photoprotection. *Photochem Photobiol Sci* 2006; **5**: 215-237.
24. Lorenzen A, Stanek C, Lang H, Andrianov V, Kalvinsh I, Schwabe U. Characterization of a G protein-coupled receptor for nicotinic acid. *Mol Pharmacol* 2001; **59**: 349-357.
25. Wise A, Foord S M, Fraser N J et al. Molecular identification of high and low affinity receptors for nicotinic acid. *J Biol Chem* 2003; **278**: 9869-9874.
26. Jacobson E L, Benavente C A, Wondrak G T, Jacobson M K. NAD and PARPs in skin cancer treatment and prevention. *Med Sci Monit* 2005; **11**: 24.
27. Kim H, Jacobson M K, Kim M, Jacobson E L, Qasem J G. A Topical Niacin Prodrug Enhances Wound Healing by Stimulation of Leptin Secretion. *J Invest Dermatol* 2002; **119**: 8405.
28. Hanley K, Jiang Y, Crumrine D et al. Activators of the nuclear hormone receptors PPARalpha and FXR accelerate the development of the fetal epidermal permeability barrier. *J Clin Invest* 1997; **100**: 705-712.
29. Sano S, Itami S, Takeda K et al. Keratinocyte-specific ablation of Stat3 exhibits impaired skin remodeling, but does not affect skin morphogenesis. *EMBO J* 1999; **18**: 4657-4668.
30. Frank S, Stallmeyer B, Kämpfer H, Kolb N, Pfeilschifter J. Leptin enhances wound re-epithelialization and constitutes a direct function of leptin in skin repair. *J Clin Invest* 2000; **106**: 501-509.
31. Bendinelli P, Maroni P, Pecori Giraldi F, Piccolini R. Leptin activates Stat3, Stat1 and AP-1 in mouse adipose tissue. *Mol Cell Endocrinol* 2000; **168**: 11-20.
32. Unger R H, Zhou Y T, Orci L. Regulation of fatty acid homeostasis in cells: novel role of leptin. *Proc Natl Acad Sci USA* 1999; **96**: 2327-2332.
33. Jacobson E L, Kim H, Kim M, Wondrak G T, Jacobson M K. Developing Topical Prodrugs for Skin Cancer Prevention. Berlin Heidelberg: Springer-Verlag, 2005.
34. Glogau R G. Aesthetic and anatomic analysis of the aging skin. *Semin Cutan Med Surg* 1996; **15**: 134-138.
35. Jacobson E L, Jacobson M K. Tissue NAD as a biochemical measure of niacin status in humans. *Methods Enzymol* 1997; **280**: 221-230.
36. Proksch E, Folster-Holst R, Jensen J M. Skin barrier function, epidermal proliferation and differentiation in eczema. *J Dermatol Sci* 2006; **43**: 159-169.
37. Lowe N, Gifford M, Tanghetti E et al. Tazarotene 0.1% cream versus tretinoin 0.05% emollient cream in the treatment of photo-damaged facial skin: a multicenter, double-blind, randomized, parallel-group study. *J Cosmet Laser Ther* 2004; **6**: 79-85.
38. Jensen J M, Folster-Holst R, Baranowsky A et al. Impaired sphingomyelinase activity and epidermal differentiation in atopic dermatitis. *J Invest Dermatol* 2004; **122**: 1423-1431.
39. Cork M J, Robinson D A, Vasilopoulos Y et al. New perspectives on epidermal barrier dysfunction in atopic dermatitis: gene-environment interactions. *J Allergy Clin Immunol* 2006; **118**: 3-21; quiz 22-23.
40. de Jongh C M, Jakasa I, Verberk M M, Kezik S. Variation in barrier impairment and inflammation of human skin as determined by sodium lauryl sulphate penetration rate. *Br J Dermatol* 2006; **154**: 651-657.
41. Kao J S, Fluhr J W, Man M Q et al. Short-term glucocorticoid treatment compromises both permeability barrier homeostasis and stratum corneum integrity: inhibition of epidermal lipid synthesis accounts for functional abnormalities. *J Invest Dermatol* 2003; **120**: 456-464.
42. Ya-Xian Z, Suetake T, Tagami H. Number of cell layers of the stratum corneum in normal skin - relationship to the anatomical location on the body, age, sex and physical parameters. *Arch Dermatol Res* 1999; **291**: 555-559.
43. Draelos Z D. Concepts in skin care maintenance. *Cutis* 2005; **76**: 19-25.
44. Draelos Z D. The effect of a daily facial cleanser for normal to oily skin on the skin barrier of subjects with acne. *Cutis* 2006; **78**: 34-40.
45. Jacobi U, Weigmann H J, Ulrich J, Sterry W, Lademann J. Estimation of the relative stratum corneum amount removed by tape stripping. *Skin Res Technol* 2005; **11**: 91-96.
46. Lee C H, Chuang H Y, Shih C C, Jong S B, Chang C H, Yu H S. Transepidermal water loss, serum IgE and beta-endorphin as important and independent biological markers for development of itch intensity in atopic dermatitis. *Br J Dermatol* 2006; **154**: 1100-1107.
47. Fluhr J W, Feingold K R, Elias P M. Transepidermal water loss reflects permeability barrier status: validation in human and rodent in vivo and ex vivo models. *Exp Dermatol* 2006; **15**: 483-492.
48. Morrow J D, Awad J A, Oates J A, Roberts Jr L J. Identification of skin as a major site of prostaglandin D2 release following oral administration of niacin in humans. *J Invest Dermatol* 1992; **98**: 812-815.
49. Carlson L A. Nicotinic acid: the broad-spectrum lipid drug. A 50th anniversary review. *J Intern Med* 2005; **258**: 94-114.
50. Benyo Z, Gille A, Bennett C L, Clausen B E, Offermanns S. Nicotinic acid-induced flushing is mediated by activation of epidermal Langerhans cells. *Mol Pharmacol* 2006; **70**: 1844-1849.

- 51 Meyers C D, Liu P, Kamanna V S, Kashyap M L. Nicotinic acid induces secretion of prostaglandin D<sub>2</sub> in human macrophages: an in vitro model of the niacin flush. *Atherosclerosis* 2006.
- 52 Monteiro-Riviere N A, Bristol D G, Manning T O, Rogers R A, Riviere J E. Interspecies and interregional analysis of the comparative histologic thickness and laser Doppler blood flow measurements at five cutaneous sites in nine species. *J Invest Dermatol* 1990; 95: 582-586.
- 53 Bhawan J, Gonzalez-Serva A, Nehal K et al. Effects of tretinoin on photodamaged skin. A histologic study. *Arch Dermatol* 1991; 127: 666-672.
- 54 Weinstein G D, Nigra T P, Pochi P E et al. Topical tretinoin for treatment of photodamaged skin. A multicenter study. *Arch Dermatol* 1991; 127: 659-665.
- 55 Tur E, Hohl D, Jetté A, Panizzon R, Frenk E. Modification of late epidermal differentiation in photoaged skin treated with topical retinoic acid cream. *Dermatology* 1995; 191: 124-128.
- 56 Bhawan J, Olsen E, Lufano L, Thorne E G, Schwab B, Gilchrist B A. Histologic evaluation of the long term effects of tretinoin on photodamaged skin. *J Dermatol Sci* 1996; 11: 177-182.
- 57 Machtinger L A, Kaidbey K, Lim J et al. Histological effects of tazarotene 0.1% cream vs. vehicle on photodamaged skin: a 6-month, multicentre, double-blind, randomized, vehicle-controlled study in patients with photodamaged facial skin. *Br J Dermatol* 2004; 151: 1245-1252.
- 58 Ghadially R, Brown B E, Sequeira-Martin S M, Feingold K R, Elias P M. The aged epidermal permeability barrier. Structural, functional, and lipid biochemical abnormalities in humans and a senescent murine model. *J Clin Invest* 1995; 95: 2281-2290.
- 59 Ghadially R, Brown B E, Hanley K, Reed J T, Feingold K R, Elias P M. Decreased epidermal lipid synthesis accounts for altered barrier function in aged mice. *J Invest Dermatol* 1996; 106: 1064-1069.
- 60 Jacobson E L, Shieh W M, Huang A C. Mapping the role of NAD metabolism in prevention and treatment of carcinogenesis. *Mol Cell Biochem* 1999; 193: 69-74.
- 61 Benavente C A, Jacobson E L. Cellular NAD status as a regulator of skin photodamage. *Proc Am Assoc Cancer Res* 2006; 47: 1438.
- 62 Ames B N. Micronutrient deficiencies. A major cause of DNA damage. *Ann N Y Acad Sci* 1999; 889: 87-106.
- 63 Jacobson-Kram D, Albertini R J, Branda R F et al. Measurement of chromosomal aberrations, sister chromatid exchange, hprt mutations, and DNA adducts in peripheral lymphocytes of human populations at increased risk for cancer. *Environ Health Perspect* 1993; 101 (Suppl. 3): 121-125.
- 64 Elias P M. Stratum corneum defensive functions: an integrated view. *J Invest Dermatol* 2005; 125: 183-200.
- 65 Lord G M, Matarese G, Howard J K, Baker R J, Bloom S R, Leclerc R I. Leptin modulates the T-cell immune response and reverses starvation-induced immunosuppression. *Nature* 1998; 394: 897-901.
- 66 Komuves L G, Hanley K, Man M Q, Elias P M, Williams M L, Feingold K R. Keratinocyte differentiation in hyperproliferative epidermis: topical application of PPARalpha activators restores tissue homeostasis. *J Invest Dermatol* 2000; 115: 361-367.
- 67 Komuves L G, Hanley K, Lefebvre A M et al. Stimulation of PPARalpha promotes epidermal keratinocyte differentiation in vivo. *J Invest Dermatol* 2000; 115: 353-360.
- 68 Draelos Z D, Ertel K, Berge C. Niacinamide-containing facial moisturizer improves skin barrier and benefits subjects with rosacea. *Cutis* 2005; 76: 135-141.
- 69 Shalita A R, Smith J G, Parish L C, Sofman M S, Chalker D K. Topical nicotinamide compared with clindamycin gel in the treatment of inflammatory acne vulgaris. *Int J Dermatol* 1995; 34: 434-437.
- 70 Wozniacka A, Wiczorkowska M, Gebicki J, Sysa-Jedrzejowska A. Topical application of 1-methylnicotinamide in the treatment of rosacea: a pilot study. *Clin Exp Dermatol* 2005; 30: 632-635.
- 71 Greatens A, Hakozaki T, Koshioffer A, et al. Effective inhibition of melanosome transfer to keratinocytes by lectins and niacinamide is reversible. *Exp Dermatol* 2005; 14: 498-508.
- 72 Blissett D L, Oblong J E, Berge C A. Niacinamide: a B vitamin that improves aging facial skin appearance. *Dermatol Surg* 2005; 860-865.
- 73 Hengge U R, Ruzicka T, Schwartz R A, Cork M J. Adverse effects of topical glucocorticosteroids. *J Am Acad Dermatol* 2006; 54: 1-15.